

Note

A two-step, regioselective synthesis of 3-hypoxanthosine and 3-hypothioxanthosine derivatives

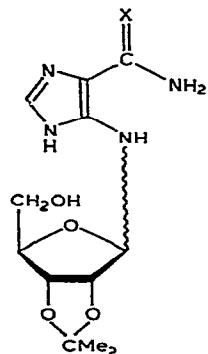
CHANTAL DE GOURCY, HAMED M. A. ABDEL-BARY, CLAUDE CHAVIS, AND JEAN-LOUIS IMBACH

Université des Sciences et Techniques du Languedoc, Laboratoire de Chimie Bio-Organique, Equipe de Recherche Associée au C.N.R.S. No. 195, Place E. Bataillon, 34060 Montpellier (France)

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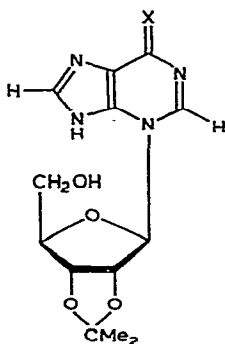
Although the 3-D-ribofuranosylpurines have interesting biological activities¹⁻⁷, their description is not well-documented, because of their relative instability^{8,9} compared to that of the 7- or 9-isomers.

Generally, by using direct-coupling techniques, the 3-D-ribofuranosylpurines can be obtained as isomeric mixtures with the corresponding 9-isomer¹⁰⁻¹⁴. However, the presence of a bulky halogen atom at position 8 of the purine ring causes pre-



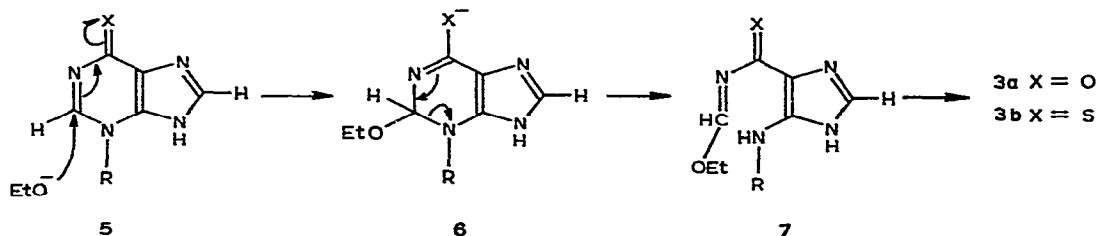
3a X = O

3b X = S



4a X = O

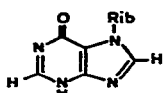
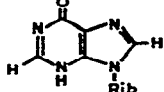
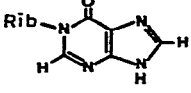
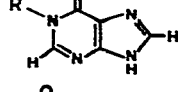
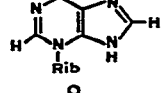
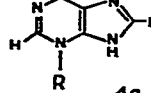
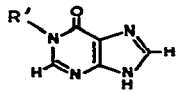
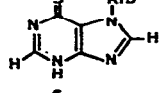
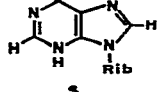
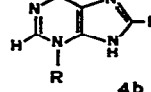
4b X = S



ponderance of N-3 substitution, but subsequent elimination of the halogen atom is not straightforward^{15,16}. Furthermore, a five-step synthesis of 3-isoxanthosine from a D-ribofuranosylpyrimidine followed by ring closure has been reported¹⁷.

TABLE I

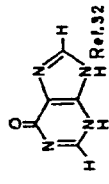
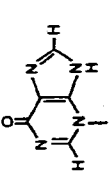
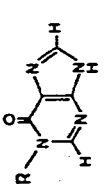
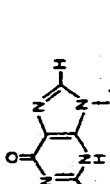
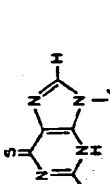
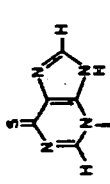
U.V. DATA: $\lambda_{\max}$ (ϵ)

	Ref.	pH 1	pH 6.7	pH 7	pH 9	pH 11	pH 13	MeOH
	25	252 (9230)		257 (8420)		263 (8830)		
	26	249 (11600)		248 (12100)		253 (13000)		
	27	249 (9580)		251 (8550)		261 (8530)		
	28	249 (9800)		250 (10000)			261 (10600)	
	16	254 (10950)		265 (13200)		270 (10900)		
		254 (11200)		263 (12300)		276 (11000)		
4a								
	28	320 (18900)		321 (23000)		321 (20800)		
	29	330 (18700)				317 (20400)		332 (16000)
	30	322 (22500)	320 (21500)				310 (22100)	
		322 (17900)		341 (22000)	339 (16000)			
4b								

^aR = 2,3-O-Isopropylidene- β -D-ribofuranosyl. R' = 2,3,5-tri-O-acetyl- β -D-ribofuranosyl.

TABLE II

¹³C-N.M.R. DATA^a

Compound ^b	C-2	C-4	C-5	C-6	C-8	C-1'	C-2'	C-3'	C-4'	C-5'		CH ₃
 4a	150.20	155.50	118.00	158.90	144.80							
	140.31	146.45	114.61	161.77	144.60	93.83	86.65	83.10	80.95	61.45	113.14	27.01 25.11
	141.13	154.16	112.90	145.38	145.38	90.95	87.44	84.56	80.76	61.25	112.89	27.01 25.16
	149.20	146.95	125.30	157.65	139.90	88.75	71.20	75.15	86.60	62.25		
	146.10	144.70	136.30	176.70	141.90	88.45	70.85	75.10	86.40	61.95		
 4b	139.09	140.59	128.45	184.74	143.74	94.41	87.73	83.93	80.65	61.25	112.89	26.87 25.01

^a The chemical shift is in CDCl₃ (internal Me₄Si). ^b R = 2,3-O-isopropylidene-β-D-ribofuranosyl. R' = β-D-ribofuranosyl.

We now describe a short, general, regioselective synthesis in the 3-D-ribofuranosylpurine series, starting from the commercially available 5-amino-4-carbamoylimidazole AICA (**1a**) or the corresponding 5-amino-4-thiocarbamoylimidazole¹⁸⁻²⁰ (**1b**).

Condensation²¹ of **1a** or **1b** with 2,3-*O*-isopropylidene-D-ribofuranose²² (**2**) gave the glycosylamines **3a** (50%) or **3b** (76%). Treatment of **3a** or **3b** in boiling ethanol with ethyl formate and sodium ethoxide afforded **4a** (50%) or **4b** (21%) after chromatography. It was observed (by t.l.c.) that, during the cyclisation step, **3a** or **3b** was always present even after prolonged boiling, presumably because of a reverse reaction (**5**→**6**→**7**), since **4a** or **4b**, when treated with sodium ethoxide in boiling ethanol, gave some **3a** or **3b**.

Mass spectrometry of **4a** and **4b** showed characteristic fragmentation processes of hypoxanthine derivatives²³. The downfield chemical-shift of the imidazole proton from δ 7.40 for H-2 in **3a** or **3b** to δ 8.25 or 8.17 for H-8 of **4a** or **4b** confirm the ring-closure process²⁴. The u.v. data in Table I show that **4a** is a 3-hypoxanthosine derivative. Since no such reference compound was available for **4b**, ¹³C-n.m.r. spectroscopy was used to confirm the site of ribosylation. Ribosylation at N-7 or N-9 of a purine ring produces³¹ a downfield shift of the carbon signal in comparison with those of the corresponding aglycon. Table II illustrates such an effect for C-2 of **4a** and 1-(2,3-*O*-isopropylidene- β -D-ribofuranosyl)hypoxanthine²⁸, as compared with hypoxanthine. The downfield shift of 7 p.p.m. for C-2 of 6-thioinosine, compared with **4b**, is consistent with a 3-D-ribofuranosyl-6-thiohypoxanthine.

The anomeric configuration of **4a** and **4b** was determined on the basis of known³³ criteria. Thus the chemical-shift difference ($\Delta\delta$ 0.15 p.p.m.) for the 2,3-*O*-isopropylidene methyl groups of a nucleoside is typical of a β anomer. Since $\Delta\delta$ is 0.24 p.p.m. for **4a** and **4b**, these compounds are 3-hypoxanthosine and 3-hypothioxanthosine derivatives, respectively.

EXPERIMENTAL

General methods. — Melting points were determined on a Gallenkamp apparatus and are uncorrected. T.l.c. was performed on aluminium foils precoated with silica gel F-254 (Merck) with detection by u.v. light or charring with sulphuric acid. Column chromatography was performed with silica gel (70–230 mesh ASTM, Merck).

Optical rotations were determined with a Perkin–Elmer 241 MC polarimeter, and u.v. spectra with an Optica model 10 spectrometer. ¹H-N.m.r. spectra were recorded with Varian T60 or HA-100 spectrometers, and ¹³C-n.m.r. spectra with Bruker WP-80 or Jeol PS-100 spectrometers on solutions in Me₂SO-*d*₆ (internal Me₄Si); chemical shifts are expressed on the δ scale. Mass spectra were recorded on a Jeol JMS D100 spectrometer by the direct-insertion procedure.

Analyses were performed by the "Service Central de Micro-Analyse du C.N.R.S.", division de Montpellier.

4-Carbamoyl-5-(2,3-O-isopropylidene-D-ribofuranosylamino)imidazole (3a). — A solution of 5-amino-4-carbamoylimidazole (**1a**; 4.21 g, 0.033 mol) in anhydrous ethanol (30 ml) containing 2,3-O-isopropylidene-D-ribofuranose (**2**) (6.36 g, 0.033 mol) and Sikkon Blue Fluka (1 g) was boiled under reflux for 30 h and then decolorised with charcoal, filtered through Celite, and concentrated *in vacuo*. The residue was purified by column chromatography (chloroform-methanol, 9:1) and crystallised from water to afford **3a** (4.95 g, 50%), m.p. 146–148°, $[\alpha]_D^{20} -6^\circ$ (*c* 1, methyl sulphoxide). P.m.r. data: δ 1.30 (s, 3 H, Me), 1.47 (s 3 H, Me), 3.30–3.50 (m, 2 H, H-5'5"), 3.70–3.94 (m, 1 H, H-4'), 4.60–4.80 (m, 2 H, H-2',3'), 5.24–5.50 and 5.68–5.90 (m, H-1', anomers), and 7.40 (s, 1 H, H-2). ^{13}C -N.m.r. data: α anomer, δ 24.85 (Me), 25.98 (Me), 62.72 (C-5'), 78.96 (C-4'), 80.97 (C-3'), 81.56 (C-2'), 84.61 (C-1'), 111.83 (CMe₂), 131.49 (C-2,5), 147.05 (C-4), and 164.30 (C=O); β anomer, δ 24.94 (Me), 26.71 (Me), 62.08 (C-5'), 78.96 (C-4'), 81.56 (C-3'), 84.24 (C-2'), 90.62 (C-1'), 112.15 (CMe₂), 131.49 (C-2,5), 146.46 (C-4), and 164.62 (C=O).

Anal. Calc. for C₁₂H₁₈N₄O₅ · H₂O: C, 45.67; H, 6.37; N, 17.71. Found: C, 45.77; H, 6.38; N, 17.72.

5-(2,3-O-Isopropylidene-D-ribofuranosylamino)-4-thiocarbamoylimidazole (3b). — A solution of 5-amino-4-thiocarbamoylimidazole^{18–20} (**1b**; 1.4 g, 0.01 mol) and 2,3-O-isopropylidene-D-ribofuranose (**2**; 1.4 g, 0.013 mol) in anhydrous ethanol (10 ml) was boiled under reflux for 3 h and then decolorised with charcoal, filtered through Celite, and concentrated. The yellow syrup crystallised on treatment with chloroform (15 ml). Recrystallisation from water afforded **3b** (2.4 g, 76%), m.p. 189–190°, $[\alpha]_D^{20} -18^\circ$ (*c* 1, methyl sulphoxide). P.m.r. data: δ 1.33 (s, 3 H, Me), 1.50 (s, 3 H, Me), 3.30–3.60 (m, 2 H, H-5'5"), 3.80–4.00 (m, 1 H, H-4'), 4.60–4.90 (m, 2 H, H-2',3'), 5.34–5.60 and 5.76–6.06 (m, 1 H, H-1'), and 7.40 (s, 1 H, H-2).

Anal. Calc. for C₁₂H₁₈N₄O₄S: C, 45.84; H, 5.77; S, 10.19. Found: C, 45.54; H, 5.44; S, 10.39.

3-(2,3-O-Isopropylidene- β -D-ribofuranosyl)hypoxanthine (4a). — To a solution of dry **3a** (5 g, 0.017 mol) in 0.7M ethanolic sodium ethoxide (110 ml) was added freshly purified ethyl formate (5.41 ml). The solution was boiled under reflux for 3 h and then decolorised with charcoal, filtered through Celite, and concentrated. The oily residue was purified by column chromatography (chloroform-methanol, 9.7:0.3) and crystallised from methanol to give **4a** (3 g, 57%), m.p. 133–135°, $[\alpha]_D^{20} -37^\circ$ (*c* 1, water). P.m.r. data: δ 1.31 (s, 3 H, Me), 1.55 (s, 3 H, Me), 3.46–3.82 (m, 2 H, H-5',5"), 4.16–4.34 (m, 1 H, H-4'), 4.98 (dd, 1 H, $J_{3',4'} 3$ Hz, H-3'), 5.30 (dd, 1 H, $J_{2',3'} 6$ Hz, H-2'), 6.18 (d, 1 H, $J_{1',2'} 2.9$ Hz, H-1'), 8.25 (s, 1 H, H-8), and 8.60 (s, 1 H, H-2). Mass spectrum: *m/e* 308 (M^+ , 18%), 293 ($\text{M}^+ - \text{Me}$, 32), 277 ($\text{M}^+ - \text{CH}_2\text{OH}$, 10), 250 ($\text{M}^+ - \text{Me} - \text{COMe}$, 14), 219 ($\text{M}^+ - \text{Me} - \text{COMe} - \text{CH}_2\text{OH}$, 60), 173 (sugar⁺, 40), 137 (base $\text{B}^+ + 2$ H, 90), 136 ($\text{B}^+ + \text{H}$, 100), 135 (B^+ , 25), 109 ($\text{B}^+ - \text{HCN}$, 60), 108 ($\text{B}^+ - \text{CO}$, 40), 81 ($\text{B}^+ - \text{HCN} - \text{CO}$ or $\text{B}^+ - \text{CO} - \text{HCN}$, 60), and 54 ($\text{B}^+ - \text{HCN} - \text{CO} - \text{HCN}$, 50).

Anal. Calc. for C₁₃H₁₆N₄O₅: C, 50.64; H, 5.23; N, 18.19. Found: C, 50.49; H, 5.18; N, 18.20.

3-(2,3-O-Isopropylidene- β -D-ribofuranosyl)-6-thiohypoxanthine (**4b**). — A solution of **3b** (1.5 g, 0.0048 mol) in ethanolic sodium ethoxide (30 ml, 0.7M) containing freshly purified ethyl formate (1.7 ml) was boiled under reflux for 3 h, and then neutralized with M HCl, treated with charcoal, filtered through Celite, and concentrated *in vacuo*. The residue was purified by column chromatography (chloroform-methanol, 8.5:1.5) and crystallised from acetone to afford **4b** (0.325 g, 21%), m.p. 205–206°, $[\alpha]_D^{20} -11^\circ$ (c 1, methanol). P.m.r. data: δ 1.32 (s, 3 H, Me), 1.56 (s, 3 H, Me), 3.45–3.84 (m, 2 H, H-5',5''), 4.16–4.42 (m, 1 H, H-4'), 4.96 (dd, 1 H, $J_{3',4'}$ 3 Hz, H-3'), 5.31 (dd, 1 H, $J_{2',3'}$ 6 Hz, H-2'), 6.25 (d, 1 H, $J_{1',2'}$ 2.5 Hz, H-1'), 8.38 (s, 1 H, H-8), and 8.67 (s, 1 H, H-2). Mass spectrum: m/e 324 (M^+ , 14%), 309 ($M^+ - \text{Me}$, 10), 293 ($M^+ - \text{CH}_2\text{OH}$, 8), 266 ($M^+ - \text{MeCOMe}$, 15), 225 ($M^+ - \text{MeCOMe} - \text{CH}_2\text{OH}$, 20), 173 (sugar $^+$, 18), 153 ($B^+ + 2 \text{ H}$, 32), 152 ($B^+ + \text{H}$, 100), 151 (B^+ , 10), 125 ($B^+ - \text{HCN}$, 25), and 114 ($B^+ - \text{CH}_2 - \text{C}=\text{S}$, 50).

Anal. Calc. for $\text{C}_{13}\text{H}_{16}\text{N}_4\text{O}_4\text{S}$: C, 48.14; H, 4.97; S, 9.88. Found: C, 48.17; H, 5.00; S, 9.65.

REFERENCES

- 1 R. FALCONER AND J. M. GULLAND, *J. Chem. Soc.*, (1939) 1369–1371.
- 2 H. S. FORREST, D. HATFIELD, AND J. M. LAGOWSKI, *J. Chem. Soc.*, (1961) 963–968.
- 3 R. LOHRMANN, J. M. LAGOWSKI, AND H. S. FORREST, *J. Chem. Soc.*, (1964) 451–459.
- 4 N. J. LEONARD AND R. A. LAURSEN, *J. Am. Chem. Soc.*, 85 (1963) 2026–2028.
- 5 K. GERZON, I. S. JOHNSON, G. B. BODER, J. C. CLINE, P. J. SIMPSON, AND C. SPETH, *Biochim. Biophys. Acta*, 119 (1966) 445–461.
- 6 A. M. MICHELSON, C. MONNY, R. A. LAURSEN, AND N. J. LEONARD, *Biochim. Biophys. Acta*, 119 (1966) 258–267.
- 7 N. J. LEONARD AND R. A. LAURSEN, *Biochemistry*, 4 (1965) 365–376.
- 8 N. J. LEONARD AND R. A. LAURSEN, *Biochemistry*, 4 (1965) 354–365.
- 9 D. B. DUNN, J. D. SMITH, AND P. F. SPAHR, *J. Mol. Biol.*, 2 (1960) 113–117.
- 10 J. W. JONES AND R. K. ROBINS, *J. Am. Chem. Soc.*, 84 (1962) 1914–1919.
- 11 H. G. FLETCHER, JR., AND C. P. J. GLAUDEMANS, *J. Org. Chem.*, 28 (1963) 3004–3006.
- 12 K. R. DARNALL AND L. B. TOWNSEND, *J. Heterocycl. Chem.*, 3 (1966) 371–373.
- 13 H. M. KISSMAN, C. PIDACKS, AND B. R. BAKER, *J. Am. Chem. Soc.*, 77 (1955) 18–24.
- 14 L. B. TOWNSEND, R. K. ROBINS, R. N. LOEPPKY, AND N. J. LEONARD, *J. Am. Chem. Soc.*, 86 (1964) 5320–5325.
- 15 C. L. SMITH AND L. B. TOWNSEND, *J. Org. Chem.*, 37 (1972) 2300–2302.
- 16 R. WOLFENDEN, T. K. SHARPLESS, N. J. LEONARD, AND I. S. RAGADE, *J. Am. Chem. Soc.*, 88 (1966) 185–186.
- 17 D. LIPKIN, C. T. CORI, AND J. A. RABI, *J. Heterocycl. Chem.*, 6 (1969) 995–996.
- 18 R. SUZUKI, T. SAITO, I. MEGURO, I. KUMASHIRO, AND T. TAKENISHI, *Jap. Pat.*, 6910 (1965); *Chem. Abstr.*, 68 (1968) 68995m.
- 19 A. YAMAZAKI, I. KUMASHIRO, T. TAKENISHI, AND M. IKEHARA, *Chem. Pharm. Bull.*, 16 (1968) 2172–2180.
- 20 C. W. SMITH, R. W. SIDWELL, R. K. ROBINS, AND R. L. TOLMAN, *J. Med. Chem.*, 15 (1975) 883–887.
- 21 G. P. ELLIS AND J. HONEYMAN, *Adv. Carbohydr. Chem.*, 10 (1955) 95–168.
- 22 G. R. BARKER AND J. W. SPOORS, *J. Chem. Soc.*, (1956) 1192–1195.
- 23 J. M. RICE AND G. O. DUDEK, *J. Am. Chem. Soc.*, 89 (1967) 2719–2725.
- 24 B. RAYNER, *Thèse Docteur ès Sciences*, Montpellier, 1977.
- 25 J. A. MONTGOMERY AND H. J. THOMAS, *J. Heterocycl. Chem.*, 5 (1968) 303–304.
- 26 J. A. MONTGOMERY AND H. J. THOMAS, *J. Org. Chem.*, 28 (1963) 2304–2310.
- 27 J. A. MONTGOMERY AND H. J. THOMAS, *J. Heterocycl. Chem.*, 5 (1968) 741–743.
- 28 A. DHAINAUT, *Thèse de 3e cycle*, Montpellier, 1975.

- 29 R. J. ROUSSEAU, R. P. PANZICA, S. M. REDDICK, R. K. ROBINS, AND L. B. TOWNSEND, *J. Org. Chem.*, 35 (1970) 631-635.
- 30 J. A. JOHNSON, H. J. THOMAS, AND H. J. SCHAEFFER, *J. Am. Chem. Soc.*, 80 (1958) 699-702.
- 31 M. T. CHENON, R. J. PUGMIRE, D. M. GRANT, R. P. PANZICA, AND L. B. TOWNSEND, *J. Am. Chem. Soc.*, 97 (1975) 4627-4636.
- 32 E. BREITMAIER AND W. VOELTER, in H. F. EBEL (Ed.), *¹³C-NMR Spectroscopy; Monographs in Modern Chemistry*, Verlag Chemie, Weinheim, 1974, p. 248.
- 33 J. L. IMBACH, J. L. BARASCUT, B. L. KAM, AND C. TAPIERO, *Tetrahedron Lett.*, (1974) 129-130.